

1. **WHEN SAMPLING, BRING ICE IN SEALED BAGS TO CHILL SAMPLES DURING SAMPLE COLLECTION.**

2. The sampler will receive the following sample kit from our lab:

Bottle Label	# of Bottles	Size	Bottle	Preservative
@UCMR3 200.8	1	500mL	Acid Rinse High-density polyethylene (HDPE) bottles with <b>white</b> polypropylene (PP) screw-caps lined with polyethylene (PE)	2mL of 18% nitric acid
@UCMR3 200.8 FB	1	250mL	Acid Rinse HDPE bottles with <b>yellow</b> PP screw-caps lined with polyethylene with <b>blue round label on bottle.</b>	1mL of 18% nitric acid

\*The sampler will receive the Field Blank (FB) filled with reagent water and preservative (**volume filled up to the bottom of the neck**). It is colored in the picture to indicate the bottle is filled (not empty) for picture purposes only.



3. Put on nitrile gloves. Open the @UCMR3 200.8 FB at the site. There is no need to keep it open for an extended period of time.



4. Reseal the bottle. Ship the FB back to the lab along with the samples.



5. If sampling from faucet, remove the aerator and screen.



6. Open the tap and let the water of the sample source run at fast flow for approximately 5 minutes.



7. Use indelible ink (pen included in kit) to clearly identify the sample bottles with the information listed below.

- Sample ID
- Source of sample, if not already on label
- Analysis required, if not already on label
- Date and Time of collection
- Preservative used, if not already on label

Client: EEA-MONROVIA Project: UCMR3 Site Name: Sample ID: Sample ID 1 Analysis: @UCMR3 200.8 Date: 03/21/14		Date: 03/21/14 Time: 10:10 Analyst: [Signature]
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8. Slow water flow to thickness of a pencil (to minimize splashing) and fill bottle.



9. Fill sample bottle up to the **shoulder**, taking care not to flush out preservatives and making sure the mouth of the bottle does not come in contact with anything other than sample water.



10. Cap and invert the bottle at least 5 times to mix the sample with the preservative.



### **SHIPPING SAMPLES AND STORAGE**

1. If shipping samples on the same day of sampling, chill samples until at or below 10°C by exchanging the ice used during sampling with available sealed bags of fresh ice.
2. **Pack chilled samples** in a cooler and add enough **FRESH** wet ice to take up 30-50% of the cooler (e.g. most of the remaining space) as recommended in our **“Wet Ice Packing Instructions.”**
3. Complete Chain of Custody during sample collection. Place completed Kit Order and completed Chain of Custody in a Ziploc bag in the cooler on top of packing material. The following information is required on the completed Chain of Custody.
  - Collector's name
  - Unique field sample ID (from UCMR database)
  - PWSID #
  - Facility ID # (from UCMR database)
  - Date and time of collection
  - Comments about the sample, if applicable
  - Sample type (EP, MR, or FB)
  - Sample Event Number (SE1, SE2, SE3, SE4)
4. **Ship via overnight service such as FEDEX, UPS, or DHL, etc.** Sample must not exceed 10°C during transit.
5. Samples **MUST** arrive at lab within 48 hours of sampling at or less than 10°C, greater than 0°C (not frozen).
6. **If samples are received more than 48 hours after sampling they must be at or less than 6°C, greater than 0°C (not frozen).**
7. If samples are received on the same day as collection, temperature may be greater than 10°C with evidence of cooling.
8. Maximum **HOLDING TIME FOR SAMPLES IS 28 DAYS** from time of collection. UCMR3 recommended hold time for sample digestates is 28 days.
9. Alternatively, cool the samples down by placing them **overnight** in a cooler with ice, or in a refrigerator (store chilled for at least 12 hours before packing for shipment). Maintain the samples cold until repacked in the cooler for shipment to the lab.

### **ADDITIONAL NOTES**

1. Do not composite (i.e., combine, mix or blend) UCMR3 samples.
2. Collect samples early enough in the day to allow adequate time to cool and to send those samples for overnight delivery to the laboratory, if not refrigerated and stored overnight before shipping.
3. Try to collect only on a Monday, Tuesday or Wednesday and ship no later than Thursday of each week, and try to **NOT** collect samples on Friday, Saturday, or Sunday unless special arrangements have been made for the receipt of samples at the laboratory within 48-hours of collection.
4. Take typical precautions when working with acids.
5. If in doubt, please review our YouTube sampling video at <http://www.youtube.com/user/EurofinsEaton>.
6. If shipping to the laboratory with **frozen gel packs** rather than wet ice, please be sure that the gel packs have **been frozen for at least 48 hours** prior to the shipment time.