

1. **WHEN SAMPLING, BRING ICE IN SEALED BAGS TO CHILL SAMPLES DURING SAMPLE COLLECTION.**

2. The sampler will receive the following sample kit from our lab:

Bottle Label	# of Bottles	Size	Bottle	Preservative
@UCMR3 522	3	125mL	Amber glass bottles with PTFE lined screw caps	6.25mg of Sodium Sulfite. 125 mg of Sodium Bisulfate, a second preservative, is attached on the side in a small vial*

*Add 125 mg of Sodium Bisulfate into the collected sample **ONLY AFTER** the Sodium Sulfite has been dissolved in the aqueous sample.

**The sampler will receive 3 sets of preserved bottles.

*** Sodium sulfite acts as a dechlorinating agent. Sodium bisulfate then acidifies the sample. The two compounds cannot be added together before sampling.



3. Put on nitrile gloves. If sampling from faucet, remove the aerator and screen.



4. Open the tap and let the water of the sample source run at fast flow for approximately 5 minutes.



5. Use indelible ink (pen included in kit) to clearly identify the sample bottles with the information listed below.

-Sample ID
 -Source of sample, if not already on label
 -Analysis required, if not already on label

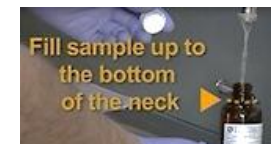
-Time and Date of collection
 -Preservative used, if not already on label



6. Slow water flow to thickness of a pencil (to minimize splashing) and fill bottle.



7. Fill sample bottle up to **bottom of the neck**, taking care not to flush out preservatives and making sure the mouth of the bottle does not come in contact with anything other than sample water.



8. Cap and invert the bottles at least 5 times or until the sodium sulfite is dissolved.



9. Open bottle and add the sodium bisulfate by shaking out from the vial.



10. After the sodium bisulfate is added, cap and invert the bottle at least 5 times or until the sodium bisulfate is dissolved.



11. Collect sample for the other 2 preserved bottles by repeating steps 6 to 10.

SHIPPING SAMPLES AND STORAGE

1. If shipping samples on the same day of sampling, chill samples until at or below 10°C by exchanging the ice used during sampling with sealed bags of fresh ice.
2. **Pack chilled samples** in a cooler and add enough **FRESH** wet ice to take up 30-50% of the cooler (e.g. most of the remaining space) as recommended in our "**Wet Ice Packing Instructions**."
3. Complete the Chain of Custody during sample collection. Place completed Kit Order and completed Chain of Custody in a ziplock bag in the cooler on top of packing material. The following information is required on the completed Chain of Custody.
 - Collector's name
 - Unique field sample ID (from UCMR database)
 - PWSID #
 - Facility ID # (from UCMR database)
 - Date and time of collection
 - Comments about the sample, if applicable
 - Sample type (EP, MR, or FB)
 - Sample Event Number (SE1, SE2, SE3, SE4)
4. **Ship via overnight service such as FEDEX, UPS, or DHL, etc.** Sample must not exceed 10°C during transit.
5. Samples **MUST** arrive at lab within 48 hours of sampling at or less than 10°C, greater than 0°C (not frozen).
6. **If samples are received more than 48 hours after sampling they must be at or less than 6°C, greater than 0°C (not frozen).**
7. If samples are received on the same day as collection, temperature may be greater than 10°C with evidence of cooling (e.g. wet ice).
8. Maximum **HOLDING TIME FOR SAMPLES IS 28 DAYS** from time of collection.
9. Alternatively, cool the samples down by placing them **overnight** in a cooler with ice, or in a refrigerator (store chilled for at least 12 hours before packing for shipment). Maintain the samples cold until repacked in the cooler for shipment to the lab.

ADDITIONAL NOTES

1. No Field Blank (FB) required.
2. Do not composite (i.e., combine, mix or blend) UCMR3 samples.
3. Collect samples early enough in the day to allow adequate time to cool and to send those samples for overnight delivery to the laboratory, if not refrigerated and stored overnight before shipping.
4. Try to collect only on a Monday, Tuesday or Wednesday and ship no later than Thursday of each week, and try to **NOT** collect samples on Friday, Saturday, or Sunday unless special arrangements have been made for the receipt of samples at the laboratory within 48-hours of collection.
5. Use appropriate caution when handling sodium bisulfate since it is highly acidic.
6. If sodium bisulfate is not added in the field the sample pH upon receipt will be too high and a replacement sample will be required.
7. If shipping to the laboratory with **frozen gel packs** rather than wet ice, please be sure that the gel packs have **been frozen for at least 48 hours** prior to the shipment time.
8. If in doubt, please review our YouTube sampling video at <http://www.youtube.com/user/EurofinsEaton>.